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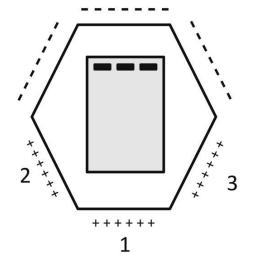
Pulse Field Gel Electrophoresis

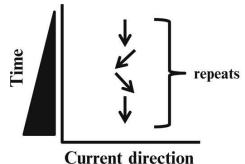
Conventional electrophoresis

2024

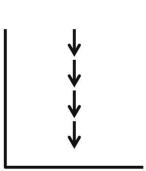
PFGE

Pulse Field Gel Electrophoresis



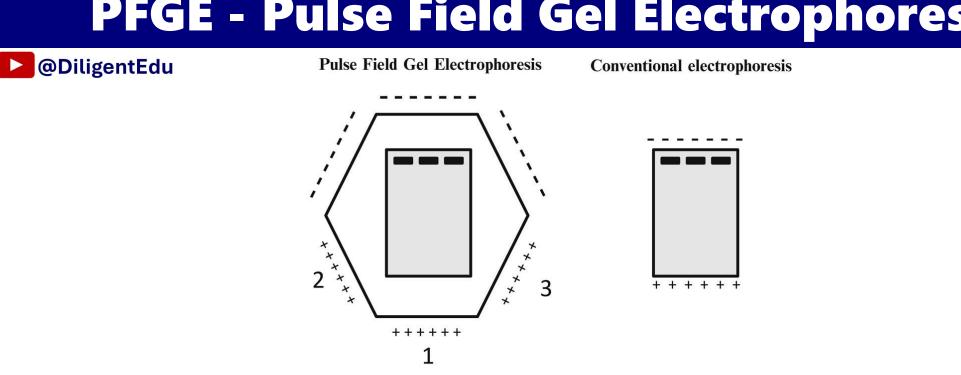


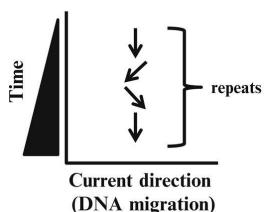
(DNA migration)

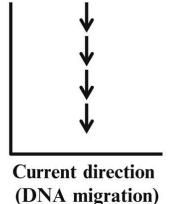


Current direction (DNA migration)











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Pulse Field Gel Electrophoresis

Pulse Field Gel Electrophoresis (PFGE) is a powerful genotyping technique used for the separation of large DNA molecules (entire genomic DNA) after digesting it with unique restriction enzymes and applying to a gel matrix under the electric field that periodically changes direction.



- Pulse Field Gel Electrophoresis
- Conventional electrophoresis uses a single electrical field to cause biomolecules to migrate through a matrix according to its mass-to-charge ratio; the migration distance of the biomolecule is indicative of its mass or size (Klotz and Zimm 1972). Conventional electrophoresis can effectively separate DNA fragments up to ~20 kb.

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- Pulse Field Gel Electrophoresis
- However, larger fragments will comigrate and appear as a large band at the top of the gel when imaged.
- In 1984, Schwartz and Cantor invented pulsed field gel electrophoresis (PFGE) to overcome this problem.



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- Pulse Field Gel Electrophoresis
- PFGE resolves DNA by alternating the electrical field between spatially distinct pairs of electrodes.
- This technique results in the separation of DNA fragments of up to ~10 Mb by their reorientation and movement at different speeds through the pores of an agarose gel.

• How Does PFGE Work?

PFGE stemmed from the observation that

- DNA molecules **elongate** upon application of an **electric field and**
- return to an **unelongated** state upon **removal** of the **electric field**;

this **relaxation rate** is dependent on the **size** of the DNA.

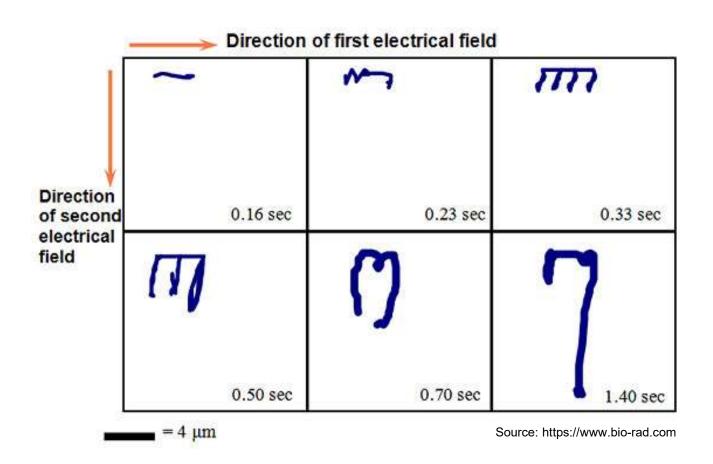


- Pulse Field Gel Electrophoresis
- When the orientation of the electric field is changed during electrophoresis, the DNA molecules must return to their elongated form prior to reorientation, thus affecting the migration rate.
- This effect can be used to greatly extend the **size** range over which electrophoretic DNA separations are possible.

- Pulse Field Gel Electrophoresis
- When the **electrical field** is applied to the gel, the DNA molecules **elongate** in the **direction of the electrical field**. The **first electrical field** is then **switched** to the **second field** according to the run specifications.
- The DNA must change conformation and reorient before it can migrate in the direction of this field.

Pulse Field Gel Electrophoresis

• As long as the alternating **fields are equal** with respect to the **voltage** and **pulse duration**, the DNA will migrate in a straight path down the gel (see below).



Time-lapse representation of DNA molecules undergoing PFGE.

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PFGE Sample Preparation

- The large size of DNA molecules to be separated by PFGE imposes certain constraints on sample preparation and handling.
- High molecular weight DNA is easily cleaved through shearing and imparts very high solution viscosity.
- For these reasons, DNA samples for PFGE are generally prepared by embedding in gel medium.



PFGE Sample Preparation

- Cellular source material is suspended in low gelling agarose and the gelled suspension is poured into molds.
- All subsequent manipulations (for example, cell lysis, protein removal, and restriction digestion) are performed by **diffusing reagents** into the resultant gel plugs.
- The processed gel plugs are then carefully loaded into wells of an agarose gel used for PFGE.



■ PFGE ... till now

- A powerful molecular typing technique
- By which genomic **DNA** is isolated from the organism of interest, followed by restriction enzyme analysis.
- The digestion products are then analyzed on an **agarose gel** by applying an **electric field** that periodically changes direction



■ PFGE ... till now

- Allowing for separation of the larger DNA fragments (entire genomic DNA) and **approximate measurement** of fragment length.
- PFGE can separate large DNA molecules (up to 10 Mb), whereas standard DNA gel electrophoresis commonly resolves fragments up to ~50 kb.
- PFGE takes 2–3 days, excluding sample preparation.

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■ PFGE ... till now

• The largest molecule so far resolved by Pulse-Field Gel Electrophoresis is estimated at **14 Mb** (Barry and Pollard, 1993)



PFGE is a variation of Agarose Gel Electrophoresis. Some of the major differences are:

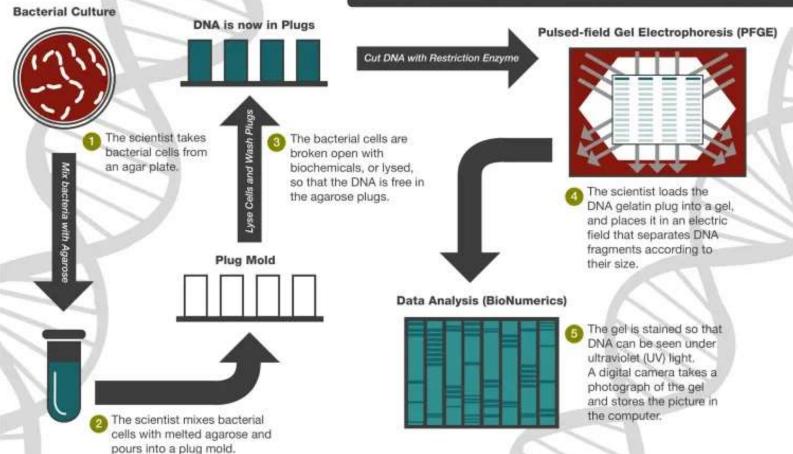
Property	Conventional gel electrophoresis	Pulsed-field gel electrophoresis	
Resolving power (Separating power)	Separates DNA fragments up to ~50 kb in size.	can separate large DNA molecules(up to 10 Mb)	
Direction of current	Current only runs in a single direction	The direction of the current switches from the primary to the secondary electrodes located at a 60° angle from the centre of the agarose gel.	
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- Steps of pulsed-field gel electrophoresis
- 1. **Preparation of Bacterial cells** Streak bacterial isolates (e.g. Staphylococcus aureus) onto TSA plates (Tryptic Soy Agar Plate) and incubate at 37 °C for 18–24 h.
- 2. Mix bacterial cells with melted agarose and pour into a plug mold Bacterial cells are embedded into an agarose gel to prevent shearing of chromosomal DNA, and DNA digestion is performed in situ.

Accessible version: https://www.cdc.gov/pulsenet/pathogens/pfge.html

sis

The Pulsed-Field Gel Electrophoresis Process



Steps of Pulsed Field Gel Electrophoresis (Source: CDC)



- Steps of pulsed-field gel electrophoresis
- Lysis of Bacterial cells The bacterial cells are broken open with biochemicals, or lysed, so that the DNA is free in the agarose plugs. The bacterial chromosome is digested using a rare cutting enzyme that recognizes specific DNA sequences ranging from 6 to 8 nucleotides, resulting in a limited number of DNA fragments of varying lengths. After DNA restriction, slices of the agarose blocks are inserted into wells of the agarose gel matrix. <u>DiligentEdu</u>©

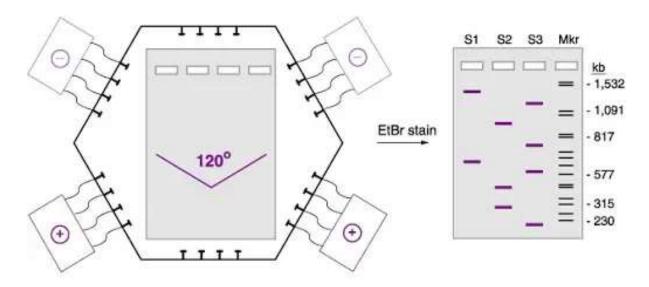
- Steps of pulsed-field gel electrophoresis
- 4. **Electrophoresis** Digested DNA samples are loaded into the DNA gelatin plug into a gel, and are subjected to separation by alternating the electric field between spatially distinct pairs of electrodes

■ PFGE – the Basic concept

Application of an **electric field** that is periodically **reorienting** causes **long DNA** molecules to **reorient** during electrophoresis. The **time required** for **reorientation** is also **inversely proportional to the size** of the DNA fragment. As **smaller molecules reorient** more **quickly** than larger molecules, separation of various DNA fragment sizes ranging from kilobases (Kb) to megabases (Mb) ensues if the gel is **run for a sufficient time**. This process generates a **fingerprint profile** that can be digitally imagined.

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This **system** consists of **24 electrodes** in an **octagon arrangement** producing a **constant electrophoresis gradient** that **switches** from the **primary to the secondary** electrodes located at a **60° angle** from the centre of the agarose gel. Consequently, this configuration causes the **DNA molecules to re-orient** in the agarose gel matrix over a **120° angle**.



Electric field alternates 120° every 90 seconds for 18 to 24 hours at 14° C



- Steps of pulsed-field gel electrophoresis
- 5. **Staining the gel** The gel is stained so that DNA can be seen under ultraviolet (UV) light.
- 6. Taking photograph and processing the Image

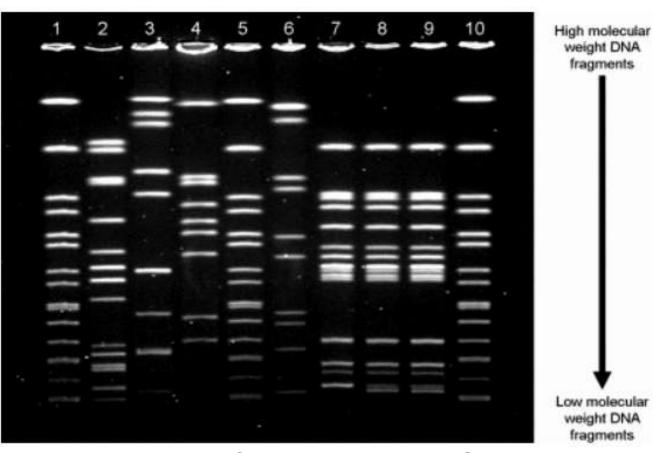
Steps of pulsed-field gel electrophoresis

Taking photograph and processing the Image

A digital camera takes a photograph of the gel and stores the picture in the computer. The obtained gel images are normalized and patterns of the DNA fragments are analyzed using various software. These software and similar other softwares help for storing, sharing and analyzing PFGE data.



Taking photograph and processing the Image







Different Types of PFGE

- CHEF (Clamped Homogeneous Electrical Field)
- PACE (Programmable Autonomously Controlled Electrodes)
- DR (Dynamic Regulation)
- FIGE (Field Inversion Gel Electrophoresis)
- AFIGE (Asymmetric Field Inversion Gel Electrophoresis)

Main Sources / Reference

- https://europepmc.org
- https://www.bio-rad.com
- https://microbeonline.com



Thank you!





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